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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/617,060	07/10/2003	Peter H. Kay	47675-52	2645
	22504 7590 05/11/2007 DAVIS WRIGHT TREMAINE, LLP			
2600 CENTURY SQUARE			GOLDBERG, JEANINE ANNE	
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

	Application No.	Applicant(s)				
Office Action Commons	10/617,060	KAY, PETER H.				
Office Action Summary	Examiner	Art Unit				
	Jeanine A. Goldberg	1634				
The MAILING DATE of this communication appears on the cover sheet with the correspondence address Period for Reply						
A SHORTENED STATUTORY PERIOD FOR REPLY WHICHEVER IS LONGER, FROM THE MAILING DATE - Extensions of time may be available under the provisions of 37 CFR 1.13 after SIX (6) MONTHS from the mailing date of this communication.  If NO period for reply is specified above, the maximum statutory period was realized to reply within the set or extended period for reply will, by statute, Any reply received by the Office later than three months after the mailing earned patent term adjustment. See 37 CFR 1.704(b).	ATE OF THIS COMMUNICATION B6(a). In no event, however, may a reply be time rill apply and will expire SIX (6) MONTHS from cause the application to become ABANDONE	N. nely filed the mailing date of this communication. D (35 U.S.C. § 133).				
Status						
1) Responsive to communication(s) filed on 13 M	<u>arch 2007</u> .					
·—	,—					
• • • • • • • • • • • • • • • • • • • •	Since this application is in condition for allowance except for formal matters, prosecution as to the merits is					
closed in accordance with the practice under Ex parte Quayle, 1935 C.D. 11, 453 O.G. 213.						
Disposition of Claims		<b>1</b> , 1				
4) Claim(s) 1-17 is/are pending in the application.	•					
4a) Of the above claim(s) is/are withdrawn from consideration.						
5) Claim(s) is/are allowed.						
6)⊠ Claim(s) <u>1-17</u> is/are rejected.						
7) Claim(s) is/are objected to.						
8) Claim(s) are subject to restriction and/or	r election requirement.					
Application Papers						
9)☐ The specification is objected to by the Examine	r.					
10)☐ The drawing(s) filed on is/are: a)☐ accepted or b)☐ objected to by the Examiner.						
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).						
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).						
11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.						
Priority under 35 U.S.C. § 119						
12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).						
a) All b) Some * c) None of:						
1. Certified copies of the priority documents have been received.						
2. Certified copies of the priority documents have been received in Application No						
3. Copies of the certified copies of the priority documents have been received in this National Stage						
application from the International Bureau (PCT Rule 17.2(a)).						
* See the attached detailed Office action for a list of the certified copies not received.						
Attachment(s)						
1) Notice of References Cited (PTO-892)	(PTO-413) ate					
<ul> <li>2) Notice of Draftsperson's Patent Drawing Review (PTO-948)</li> <li>3) Information Disclosure Statement(s) (PTO/SB/08)</li> </ul>	5) 🔲 Notice of Informal F					
Paper No(s)/Mail Date 3/07. 6) Other:						

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#### **DETAILED ACTION**

1. This action is in response to the papers filed March 13, 2007. Currently, claims1-17 are pending.

- 2. A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on March 13, 2007 has been entered.
- 3. All arguments have been thoroughly reviewed but are deemed non-persuasive for the reasons which follow.
- 4. Any objections and rejections not reiterated below are hereby withdrawn.

## **New Grounds of Rejection**

#### Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

5. Claims 1-6, 14-17 are rejected under 35 U.S.C. 103(a) as being unpatentable over Elsas, II et al. (US Pat. 6,207,387, March 27, 2001) in view of either Kool (US Pat.

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4. 4004

5,808,036, September 1998) or Kool (US Pat. 5,426,180, June 1995) and in further view of either Tyagi et al (US Pat. 6,150,097, November 2000) or Coull et al (US Pat. 6,355,421, March 2002).

Elsas, II et al. (US Pat. 6,207,387, March 27, 2001) teaches detecting mutations in genes by determining the melting temperature of the hybrid of the amplified DNA and the specific oligonucleotide (col. 10, lines 20-25). Elsas teaches that "under identical conditions, two strands that are not exactly complementary, differing by even one nucleotide, will be less stable and will dissociate at a temperature which exactly complementary hybrids remain paired (col. 8, lines 50-57). Elsas teaches that the melting temperature between a mismatched hybrid will denature at a lower temperature than a exact matched hybrid (col. 10, lines 37-40). Elsas also teaches that fluorescence energy transfer is a specific application of this approach (col. 10, lines 49-50). Elsas teaches the different melting temperatures for the allele specific probe and detection of fluorescence (col. 11).

While Elsas teaches detecting different nucleic acids based upon melting temperature, Elsas does not specifically teach the structure of fluorescence energy transfer and does not teach using the fluorescence energy transfer for detecting methylation.

Kool '036 teaches "it has been known for some time that methylation at the C-5 position of cytosine, forming the naturally-occurring base m5 C, raises the Tm of duplex DNA in which it occurs, relative to unmethylated sequences (Zmudzka et al., 1969, Biochemistry 8:3049). In order to investigate whether addition of this methyl group

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would stabilize stem-loop:target complexes, the melting temperatures of methylated and unmethylated stem-loop oligonucleotides are compared. Use of the natural base m5 in place of C can increase stability substantially.

Kool '180 teaches "it has been known for some time that methylation at the C-5 position of cytosine, forming the naturally-occurring base m.sup.5 C, raises the Tm of duplex DNA in which it occurs, relative to unmethylated sequences (Zmudzka et al., 1969, Biochemistry 8: 3049). In order to investigate whether addition of this methyl group would stabilize circle:target complexes, two analogs of circle 7 (having SEQ ID NO: 6) were synthesized. In one circle, the six C's in the binding domains were methylated leaving the loop unmethylated (Me6). In the second circle, all twleve C's were methylated (Me12). Melting temperatures for the complexes of these methylated circle with target 5 were measured. The Me.sub.6 complex had a Tm of 71.1 degree C. (compared to 61.8 degree C. for the unmethylated circle), and the Me.sub.12 circle had a Tm of 72.4 degree C. Thus, use of the natural base m5 C in place of C increased stability substantially, and in one case resulted in a 12-base complex which melted 10.6 degree C. higher than an unmethylated circle and 28.6 degree C. higher than the corresponding unmethylated Watson-Crick duplex.

Moreover, Tyagi et al. (herein referred to as Tyagi) teaches using nucleic acid hybridization probes having a first conformation when not interacting with a target and a second conformation when interacting with a target and having the ability to bring a label pair into touching contact in one formation but not the other (abstract). Tyagi teaches using quenching molecules and other fluorophores as efficient quenching

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moieties for fluorophores when attached to nucleic acid hybridization probes (col 3, lines 40-43)(limitations of Claim 2). The probes of Tyagi contain a hairpin structure which comprise single stranded loop of the hairpin and two arm sequences which form a double stranded stem hybrid (col. 5, lines 10-15). Tyagi teaches that the molecular beacon probes may have target recognition sequences 7-140 nucleotides in length (col 5, lines 24-25)(limitations of Claims 3-6). Additionally Tyagi teaches the arms that form a stem hybrid or stem duplex are 3-25 nucleotides in length (col. 5, lines 26-27)(limitations of Claim 16). Tyagi teaches a kit which contains a hairpin probe with labels (col 19, lines 66-67, Claim 12)(limitations of Claim 18).

Coull et al. (herein referred to as Coull) teaches methods of detecting target sequences using a probe which has a measurable change in one property of at least one donor or acceptor moiety of the probe which can be used to detect, identify or quantitated the target sequence in a sample. As seen in Figure 11, configuration III, a probing segment is flanked on either side by a arm segment and either a quencher and fluorophore. The hairpin loop and stem structure allows energy transfer between donor and acceptor moieties linked at opposite ends of the nucleic acid polymer (col. 7, lines 30-37). The probing segments is designed to hybridized to at least a portion of a target sequence (col. 8, lines 35-36). In the method of Coull, a sample is contacted with the molecular beacon and a change in detectable signal associated with at least one donor or acceptor moiety of the probe is detected, identified or quantitated. Coull teaches that the assay may be used to detect a target sequence which is specific for a genetically based disease including cancer. Coull teaches that the probing sequence hybridizes to

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the entire target sequence (col. 16, lines 20-25). The probing sequence will generally have a length between 5-30 units in length (col. 16, lines 32-34)(limitations of Claims 3-6). The arm segments are 2-6 units in length (limitations of Claim 16). Coull teaches that shorter probes are less costly to synthesize, are generally easier to purify and should exhibit few non-specific interactions since they will comprise less nucleobase sequence diversity (col. 20, lines 29-32)(limitations of Claim 15). Coull teaches kits which comprise one or more PNA Molecular Beacons (col. 24, lines 51-67)(limitations of Claim 18). Coull teaches considerable analysis of the Tm melting temperature for the stem-loop hairpin probes (col. 20, 37-44). Coull teaches that the probes exhibit a low inherent noise (background) and an increase in detectable signal upon binding of the probe to a target sequence (col. 7, lines 40-42). Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Elsas for detecting different nucleic acids based upon different hybridization properties including melting temperatures with the teachings of either Tyagi or Coull which discuss and describe stem-loop and fluorescence energy transfer with the teachings of either Kool '036 or Kool '180 teaching the increased stability of methylated DNA. Detecting nucleic acids based upon different melting temperatures and dissociation properties was used to identify mismatches in nucleic acids. Both Kool '036 and Kool '180 teach that methylated DNA and unmethylated DNA have different melting temperatures. Kool '180 teaches "the use of the natural base m5 C in place of C increased stability substantially." Moreover, Kool '180 specifically provides analysis of a small region with 0, 6 or 12 methylated C's and illustrates a drastic difference in melting temperature.

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The ordinary artisan would have recognized based upon the teachings that in the art, namely Kool '036 and Kool '180, that in addition to mismatched DNA, methylation could also be detected based upon different melting temperature and dissociation rates. Combining the teachings of Elsas and either Kool '036 or Kool '180 would yield an assay which would detect methylated nucleic acids as compared to unmethylated nucleic acids. Detection of methylated nucleic acids as compared to unmethylated nucleic acids is of interest to the clinical diagnostics because numerous genes are methylated in cancer as compared to unmethylated in normal tissue. Therefore, combining the teachings of Elsas and Kool '036 or Kool '180, a hybridization assay for differentiation methylated DNA from unmethylated DNA does not require the use of enzymes, solid supports would facilitate the detection of methylation in genes and may be used as an indicator for cancer. Moreover, the use of hairpin stem-loop probes with fluorophores and quenchers for detecting target nucleic acids in samples is taught by both Tyagi and Coull would have provided a fluorescent detection assay which is easily detectable in a single tube which does not require subjection to a gel or solid support. The use of FRET allows for the direct detection of nucleic acid target sequences without the requirement that labeled nucleic acid hybridization probes or primers be separated from the hybridization complex prior to detection (Coull et al. col. 1, lines 45-50). Therefore, using the specific teachings about fluorescence energy transfer techniques, as described in Coull and Tyagi, would facilitate the fluorescence energy transfer method for detecting different nucleic acids as taught by Elsas. Therefore, given all of the teachings well known in the art, at the time the invention was made, a FRET-like

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method for detection of different nucleic acids based upon the know property that methylated and unmethylated DNA molecules have different hybridization properties would have been obvious to the ordinary artisan.

6. Claims 7, 10, 11, 12, 13 are rejected under 35 U.S.C. 103(a) as being unpatentable over Elsas, II et al. (US Pat. 6,207,387, March 27, 2001) in view of either Kool (US Pat. 5,808,036, September 1998) or Kool (US Pat. 5,426,180, June 1995) and in further view of either Tyagi et al (US Pat. 6,150,097, November 2000) or Coull et al (US Pat. 6,355,421, March 2002) as applied to Claims 1-6, 14-17 above, and further in view of Herman et al. (US Pat. 6,265,171, July 2001).

The combination of Elsas, Kool '036 or Kool '180 and Tyagi or Coull does not specifically teach detecting methylation in GSTpi or calcitonin which is differentially expressed in cancer versus a normal state.

However, Herman et al. (herein referred to as Herman) teaches numerous genes which are differentially methylated at CpG islands in neoplastic versus normal tissue (limitations of Claim 7). These genes include GSTpi and calcitonin (limitations of Claims 10, 12). Herman also teaches that CpG island differential methylation may be detected in prostate cancer (col. 112, Claim 12)(limitations of Claim 12). Aberrant methylation in the 5' promoter of E-cadherin is prostate, breast and many other carcinomas (col. 27, lines 5-10).

Therefore, using the method of Elsas, Kool '036 or Kool '180 and Tyagi or Coull in view of the teachings of differential methylation in gluthathione-S-transferase-II(pi)

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and calcitonin. The ordinary artisan would have been motivated to have detected methylation in these two specific genes because Herman teaches that they contain methylated CpG neoplastic versus normal tissue.

7. Claims 7, 8, 9 are rejected under 35 U.S.C. 103(a) as being unpatentable over Elsas, II et al. (US Pat. 6,207,387, March 27, 2001) in view of either Kool (US Pat. 5,808,036, September 1998) or Kool (US Pat. 5,426,180, June 1995) and in further view of either Tyagi et al (US Pat. 6,150,097, November 2000) or Coull et al (US Pat. 6,355,421, March 2002) as applied to Claims 1-6, 14-17 above, and further in view of Kay et al (Leukemia and Lymphoma, Vol. 24, pages 211-220, 1997).

The combination of Elsas, Kool '036 or Kool '180 and Tyagi or Coull does not specifically teach detecting methylation in Myf-3 which is differentially expressed in cancer versus a normal state.

However, Kay et al. (herein referred to as Kay) teaches the Myf-3 gene is abnormally hypermethylated in non-Hodgkins lymphoma (abstract).

Therefore, using the method of Elsas, Kool '036 or Kool '180 and Tyagi or Coull in view of the teachings of differential methylation in gluthathione-S-transferase-II(pi) and calcitonin. The ordinary artisan would have been motivated to have detected methylation in these two specific genes because Herman teaches that they contain methylated CpG neoplastic versus normal tissue.

### **Conclusion**

#### 8. No claims allowable.

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9. Any inquiry concerning this communication or earlier communications from the examiner should be directed to examiner Jeanine Goldberg whose telephone number is (571) 272-0743. The examiner can normally be reached Monday-Friday from 7:00 a.m. to 4:00 p.m.

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If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Ram Shukla, can be reached on (571) 272-0735.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

The Central Fax Number for official correspondence is (571) 273-8300.

Jeanine Goldberg Primary Examiner May 8, 2007